LEPIDIUM SATIVUM L. AS TEST-ORGANISM FOR ASSESSMENT OF ENVIRONMENTAL POLLUTION

Danute Marčiulionienė¹ Danguole Montvydienė¹ Vilmante Karlavičienė^{2,3} William Hogland³ ¹Institute of Botany, Lithuania ²Vilnius Gediminas Technical University, Lithuania ³University of Kalmar, Sweden

ABSTRACT

Polluting substances when released into environment become one of the main factors of environmental pollution since they cause changes to the chemical composition of water, bottom sediments and soil; they also disturb the biological balance and self-cleaning processes of the ecosystem that may lead to unpredictable alterations within the ecosystem. Together with the chemical and physical techniques the methods of bioindication and biotesting are employed in the current research on environmental pollution and its impact on the ecological integrity of ecosystems.

The aim of this paper was to determine the sensitivity of a test-organism L. sativum to the toxic impact of different types of samples (liquid or solid: surface water, industrial wastewater, industrial storm water, landfill leachate, lake bottom sediments and sludge from industrial storm water sedimentation tank) from urban environment and chemical substances and to demonstrate the possibilities of application of this plant in both the toxicological investigation and environmental pollution assessment.

The results of conducted investigation have proved that L. sativum is a sufficiently universal, cheap and sensitive biotest for determination of toxicity of different types of samples (storm water, bottom sediments, soil, wastewater from industrial sites, landfill leachate) and included chemical substances. Therefore, it can be successfully applied in the environmental pollution assessment.

KEYWORDS

Lepidium sativum L.; Toxicity; Environmental pollution; Surface water; Storm water; Landfill leachate; Bottom sediments, Environmental pollution assessment

1 INTRODUCTION

Polluting substances when released into environment become one of the main factors of environmental pollution since they cause changes to the chemical composition of water,

Kalmar ECO-TECH '05 and The Second Baltic Symposium on Environmental Chemistry KALMAR, SWEDEN, November 28-30, 2005

bottom sediments and soil; they also disturb the biological balance and self-cleaning processes of the ecosystem that may lead to unpredictable alterations within the ecosystem.

Most frequently the pollution (or quality) of water, bottom sediments and soil is assessed using physical and chemical methods; however, these methods have some drawbacks, for instance a) they do not reflect the transformation of polluting substances in water, their interaction and bioavailability; b) they do not allow assessing the integrated toxic impact of polluting substances on organisms since environmental pollution usually tends to be multicomponent; c) the concentration of polluting substances present in soil or water does not always indicate the toxic impact of these substances on organisms since the toxicity is defined as a biological feature [1, 2]. Together with the chemical and physical techniques the methods of bioindication and biotesting are employed in the current research on environmental pollution and its impact on the ecological integrity of ecosystems [3].

Natural water and wastewater might be toxic, despite of the fact that it is lightly polluted [4]. Therefore, it is essential to know not only the concentration of polluting substances in water but also the biological and toxic effects caused by them, which may be only assessed applying the set of biotests. The main objective of biotests' application is the effective determination of toxicity level of polluting substances, wastewater and surface water, bottom sediments and soil in order to prevent the release of polluting substances into environment, to increase pollution control, to guarantee the efficiency of wastewater treatment for maintenance of natural water quality [5].

With the employment of test-organisms pollution impact is determined by the reactions of the organisms; i.e. the pollution impact on organisms is assessed. *Lepidium sativum L.* (a garden cress) classified within the *Brassicaceae* family is widely applied for biotesting in Canada, the USA and the EU countries (OECD 208, 2003; Phytotoxkit). The impact of tested samples on *L. sativum* may be evaluated by different test-parameters, for example, a number of germinated seeds, length of roots as well as the biomass of the whole plant or separate parts of it.

The aim of this paper is to determine the sensitivity of a test-organism L. sativum to the toxic impact of different types of samples (liquid or solid: surface water, industrial wastewater, industrial storm water, municipal landfill leachate, lake bottom sediments and sludge from storm water sedimentation tank) from urban environment and chemical substances and to demonstrate the possibilities of application of this plant in both the toxicological investigation and environmental pollution assessment.

2 OBJECTS AND METHODS

The substances investigated include: water and bottom sediments of different pollution levels from wastewater canals of Ignalina Nuclear Power Plant (INPP), water and bottom sediments samples from the monitoring stations of Lake Drūkšiai (Ignalina NPP cooler) located closer or further to the Plant, storm wastewater and sludge from the scrap metal processing company (Sweden), leachate water from a landfill (Lithuania), water from reclamation ditches, fuel oil, heavy metals (HM) and their mixtures.

The sludge formed in the storm water sedimentation tank at the scrap metal processing company was 10, 20, 40, 60 and 100 times diluted with distilled water, i.e. until the impact indicator did not statistically differ from the control. In investigation of fuel oil impact on L.

sativum 0.1g/L; 1.0 g/L; 5.0 g/L; 10.0 g/L; 25.0 g/L; 50.0 g/L and 100.0 g/L concentrations of fuel oil were applied.

In investigation of heavy metals impact (concentrations range from 0.01 to 100 mg/L) on *L.* sativum the following chemically pure salts were used: $Cd(NO_3)_2 \cdot 4H_2O$, $CuSO_4 \cdot 5H_2O$, $K_2Cr_2O_7$, $MnCl_2 \cdot 4H_2O$, $Pb(NO_3)_2$, $ZnSO_4 \cdot 7H_2O$ (Reachim, St. Petersburg, Russia) and Ni(NO₃)₂, (Merck, Darmstadt, Germany).

The composition of heavy metal mixture (HMM) was based on the average data of the annual amounts of HM in wastewater (WW) discharge from Ignalina Nuclear Power Plant into Lake Drūkšiai in 1996. The stock solution of HMM (1000 times higher than WW of INPP) for better solubility of HM salts was prepared in acidified distilled water using the chemically pure substances such as $Cd(CH_3COO)_2 \cdot 2H_2O$, $K_2Cr_2O_7$, $CuSO_4 \cdot 5H_2O$, $NiSO_4 \cdot 7H_2O$, $MnSO_4 \cdot 5H_2O$, $Pb(NO_3)_2$, and $ZnSO_4 \cdot 7H_2O$ (Reachim, St. Petersburg, Russia). The basic heavy metal mixture solution (HMM₁₀₀) was obtained by diluting stock solution for 1000 times to acquire HM concentrations in WW of INPP (in mg/L): Pb – 0.0142; Cd – 0.000018; Cr – 0.00028; Ni – 0.00021; Cu – 0.0075; Mn – 0.0099, and Zn – 0.064, respectively. The following mixture concentrations were applied: HMM₁₀, HMM₂₅₀, HMM₅₀, HMM₁₀₀, HMM₂₅₀₀. The distilled water was used for dilution. The test solutions initially were adjusted to pH=5.5±0.2 with 0.1 M HCl or NaOH.

The test with *L. sativum* was carried out following a modified Magone (1989) method [6]. In brief, 9 ml of distilled water (as a control) or testing sample solution was pipetted onto three layers of filter paper fitted into a 9-cm glass Petri dish. The layers of 5 to 6 cm of storm water and sludge from the storm water sedimentation tank located at the scrap metal processing company was spread over the Petri dishes. Twenty-five healthy looking and similar size seeds of *L. sativum* were distributed evenly on the filter paper. The Petri dishes were placed in the dark room at $24\pm1^{\circ}$ C for 48 hours. Afterwards the seed germination and root length of seedlings were measured. Germination of seeds and length of *L. sativum* roots in distilled water were 96 ± 4% and from 25.6±0.7 to 36.0±2.8 mm respectively. The experimental set of each testing scheme involved 3 control dishes and 3 or 5 replicates for each tested sample.

The level of toxic impact on *L. sativum* was assessed by the modified method of [7]. According to the causing percent of root growth inhibitions of 100-60%, 61-40%, 41-20% and lower than 19% the toxic impact of tested sample solutions or bottom sediments on *L. sativum* was classified as very strong, strong, moderate and weak, respectively. The tested sample was non-toxic if the indicator of root growth of *L. sativum* did not statistically differ from the control indicator, and was extremely toxic if the seed did not germinate at all.

The standard error (SE) was calculated for the statistical evaluation of data. In determining whether the results were statistically different from the control indicators the set of two-intake comparison tests of *Statgrahics plus Version 2.1.* program when p<0,05 was applied.

The 50% effective concentration of a toxicant (EC₅₀) is one of the most important parameters for toxicity assessment calculated by the regression curve "concentration-response", lgL=a+b·lgEC, when L stands for the root length in‰; a and b are constants and EC is the effective toxicant concentration. EC₅₀ is the toxicant concentration leading to the 50% reaction of test-organisms when the important alteration of life activity other than mortality is chosen. The EC₅₀ of 24, 48 or 96 hours or longer is subject to the duration of impact. In case of *L. sativum* a 48 hour EC₅₀ (48h-EC₅₀) was applied.

3 RESULTS AND DISCUSSION

The investigation of Ignalina NPP wastewater impact on *L. sativum* (from 1996 to 2000) indicated that in most cases the number of germinated seeds did not differentiate (p<0,05) or almost did not differentiate from the control indicators. However, the growth rates of *L. sativum* roots due to the impact of the analysed wastewater ranged from 54% to 95% (Fig. 1).

From the total amount of tested wastewater from Ignalina NPP during 1996–2000 only the results of year 1998 were positive. In most cases wastewater from WWTP was the most toxic; the growth rates of roots there ranged from 54% to 65%. The water of Lake Drūkšiai was also toxic and made impact on the growth of *L. sativum* roots since the growth rates (except in 1998) ranged from 55% to 84% (see *Figure 1*). The rate of toxicity of Lake Drūkšiai water was dependent on the pollution level of Ignalina NPP wastewater, distance from the pollution resource, wind direction, water flow and mixing of water masses.

The toxicity investigation of both Lake Drūkšiai surface water and bottom sediments as the location of concentration of various toxic substances was executed in order to assess the ecotoxical state of the lake. The impact of bottom sediments samples collected from 132 locations of the Lake Drūkšiai on *L. sativum* was assessed for the evaluation of ecological situation (see *Figure 2*).

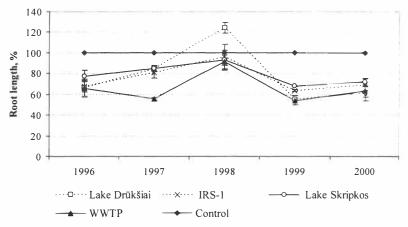


Figure 1. Toxic effect of Ignalina NPP wastewater and water of Lake Drūkšiai, the cooler of Ignalina NPP, on L. sativum;

IRS-1 – industrial wastewater and storm water run-off canal; WWTP – wastewater treatment plant; Lake Skripkos – the lake which is on the route of the wastewater from WWTP to Lake $Dr\bar{u}k$ šiai.

The results indicated that only few samples of tested bottom sediments were non-toxic (see Fig. 2). A very strong toxic impact was indicated by 13 samples of bottom sediments collected from eutrophicated shoal zones of the lake where the municipal wastewater was disposed as well as the littoral areas of the lake. The results of samples of bottom sediments discharged from the main, very deep and windy area of the lake with the industrial and storm water as well as heated wastewater disposed there indicated a moderate toxic impact.

Kalmar ECO-TECH '05 and The Second Baltic Symposium on Environmental Chemistry KALMAR, SWEDEN, November 28-30, 2005

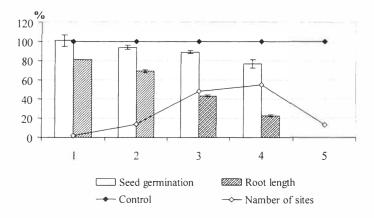


Figure 2. Toxic effect of Lake Drūkšiai bottom sediments on L. sativum 1. Non toxic or weak toxic effect; 2. Medium toxic effect; 3. Strong toxic effect; 4. Very strong toxic effect; 5. Extremely toxic effect.

It is quite possible that the toxic substances disposed into Lake Drūkšiai together with the wastewater of Ingalina NPP were rapidly drifted to the littoral areas of the lake due to the intensive mixing and strong water currents produced in the intake and heated water canals. The results of the highest toxicity of Lake Drūkšiai bottom sediments in most cases coincided with the borders of a area exposed to the highest level of geochemical contamination identified by K. Jokšas and others in 1997 [8]. The data obtained identified bottom sediments as more informative type of sample for the assessment of watersheds pollution than water.

The research results of toxic impact of storm water and sludge from the scrap metal processing plant on *L. sativum* indicated that the storm water was slightly toxic and sludge was very toxic (see Fig. 3). After sludge had been 10 and 20 times diluted with distilled water it became of moderate toxicity, meanwhile 40 and 60 times diluted it was slightly toxic. The impact of sludge diluted for 100 times to this plant was not statistically different from the control indicators (see *Figure 3*). Depending on the sludge dilution the growth rate of *L. sativum* roots ranged from 56 to 99 %, and the germination rate of seeds from 89 to 96 % (see *Figure 3*).

The results of toxicity investigation of leachate water from a municipal landfill (Lithuania) rich with industrial waste, water from reclamation ditches where leachate was disposed and pond water where water from ditches was disposed to *L. sativum* indicated that all investigated watersheds were toxic; however, the level of toxicity differed (see *Figure 4*).

Leachate water from the landfill and reclamation ditch (No.2) closest to the leachate tank was very toxic since the germination of seeds equalled to zero. The toxicity of water from reclamation ditches No. 3 and No. 4 was also very strong and strong respectively, although they were situated further from the leachate tank. In comparison with the control indicators the germination rates were 20% and 50% respectively, and the root growth was 36% and 54% respectively (see *Figure 4*).

Kalmar ECO-TECH '05 and The Second Baltic Symposium on Environmental Chemistry KALMAR, SWEDEN, November 28-30, 2005

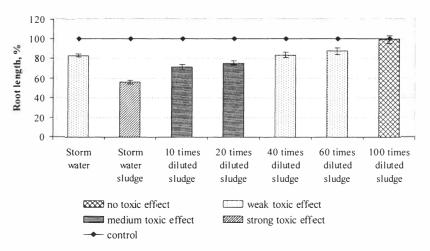


Figure 3. Toxic effect of industrial storm water and sludge(from storm water sedimentation tank) formed at metal scrap processing industry (Sweden) area on L. sativum

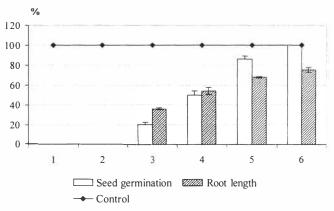


Figure 4. Toxic effect of leachate from municipal landfill rich with industrial waste (Lithuania) on L. sativum (samples were taken from leachate tank, reclamation ditches, pond and out-flow ditch)

1 – leachate tank; 2–4 – reclamation ditches to which leachate is discharged; 5 – pond to which water from reclamation ditches is collected; 6 – rivulet running from the pond.

Water from reclamation ditch (No.4) floated into the pond (No.5), which was of moderate toxicity, since 86% of *L. sativum* seeds germinated in this water and the length of roots reached 68% (see Figure 4). 100% of seeds germinated from the water of rivulet (No. 6) running from the pond; however, the length of roots did not exceed 70% in comparison with the control indicator (see Figure 4). The water was also of moderate toxicity.

The investigation of toxic impact of fuel oil, heavy metals and their mixture on germination of seeds and root growth of *L. sativum* was carried out in order to evaluate the sensitivity of *L. sativum* to separate toxicants included into the composition of wastewater produced by industrial plants and landfill leachate. Also the 50% effective concentration of these toxicants (48h-EC₅₀) was applied allowing the comparison of the impact of toxicants on different organisms.

The results of the investigation indicated that the indicators of L. sativum seed germination in all tested oil concentrations did not differ from the control indicators. Also the results showed

that the growth rates of roots in the minimal oil concentration (0.1 g/L) did not statistically differ from the control indicators. With the oil concentration of 1 g/L the length of roots was 84.2%; meanwhile with the oil concentration of 5 g/l the length reached 54.4% (see *Figure 5*).

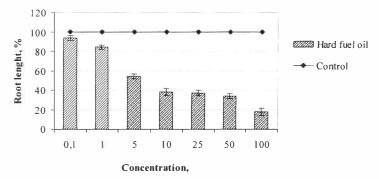


Figure 5. Toxic effect of hard fuel oil on L. sativum

With the oil concentration of 10 –100 g/l the growth of *L. sativum* roots decreased from 38.2 to 17.9% respectively (see *Figure 5*). It was calculated that the 50 % effective fuel oil concentration (48-h EC₅₀) according to the growth rates of *L. sativum* roots was 8.7 g/L. Meanwhile, 14 day-EC₅₀ for *Spirodela polyrrhiza* (L.) classified within the *Lemnaceae* family was 18.7 g/L; 96h-EC₅₀ of fuel oil for the adults' rainbow trout (*Oncorhynchus mykyss* Wallbaum.) made 3.0 g/L [9].

Investigation of the toxicity of heavy metals (Cd, Cu, Cr(VI), Mn(II), Pb, and Zn) on *L.* sativum showed that seed germination in the tested HM concentrations differed insignificantly from the control indicator, except, with the highest concentration (10 mg/L) of Cu, which decreased seed germination by 20%. The toxicity of HM was assessed using the 48-EC₅₀ values (i.e. metal concentration that induce 50% root growth inhibition of *L.* sativum as percentage of control indicator, in 48-h experiments). The comparison of the 48-EC₅₀ values of all tested HM showed that Cr (1.8 mg/L) was most toxic to *L.* sativum and Mn(II) was the least toxic (see *Figure 6*). It was estimated that the toxicity of the test metals decreased in the following order Cr ∞ Cu > Cd > Ni = Pb > Zn > Mn (*Figure 6*). Very few literature data was available for comparison with our results. Wang (1986) found that the order of toxicity for root growth of lettuce and millet was the following: Ni > Cd > Cu > Cr(VI) > Zn > Mn and Cu, Ni > Cd > Cr(VI) > Zn > Mn, respectively [10]. In 1998 Fargasova presented the following order of metals toxicity for root elongation of *Sinapis alba*: Cu²⁺ > MoO₄²⁻ > Ni²⁺ > Mn²⁺ > VO₄³⁻ [11].

Kalmar ECO-TECH '05 and The Second Baltic Symposium on Environmental Chemistry KALMAR, SWEDEN, November 28-30, 2005

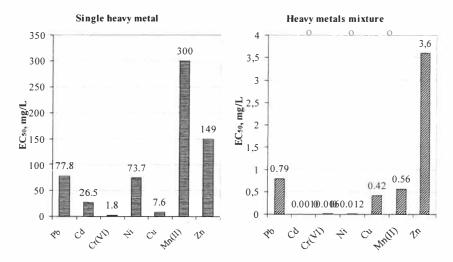


Figure 6. 50% effective concentrations (48 $h - EC_{50}$, mg/L) of single heavy metals and their in the heavy metals mixture for L sativum

The investigation of the toxic impact of heavy metal mixture on *L. sativum* identified that concentrations of HMM₁₀, HMM₂₅, HMM₅₀, HMM₁₀₀, HMM₂₅₀ and HMM₅₀₀ had no impact on the germination of seeds and root growth of this plant (p<0.05). The indicators of seed germination of M₂₅₀₀ and HMM₅₀₀₀ concentrations did not differ form the control (p<0.05), however, the root growth rate decreased to 78.2 and 56.3% respectively. The calculation of 48h-EC₅₀ values of the HM mixture for tested plant showed that the root length decreased by 50% when the basic concentrations of HM in the mixture was increased to 56.2 times. It was established that the concentrations of HM in the mixture that caused the analogous toxic effect (48h-EC₅₀) on *L. sativum* were from several to hundred times lower than the concentration of single metals that caused the same effect. For example, 48h-EC50 values of single Cu and Ni for *L. sativum* were 7.6 and 73.7 mg/L, while the concentrations of these metals in the mixture was only 0.4 and 0.012 mg/L, respectively (see *Figure 6*). It shows that the toxicity of HMM can be stipulated by the interaction within the mixture.

The results of conducted investigation have proved that *L. sativum* is a sufficiently universal, cheap and sensitive biotest for determination of toxicity of various liquid or solid samples, for example: storm water, bottom sediments from surface water, soil, wastewater from industrial sites, landfill leachate and included chemical substances. Therefore, it can be successfully applied in the environmental pollution assessment.

REFERENCES

- [1] Wundram, M, Selmar D, Bahadir, M., 1997. Representative evaluation of phytotoxicity - reliability and peculiarities. *Angew Bot* 71, 139-143.
- [2] Marčiulionienė, D, Montvydienė, D, Kazlauskiene, N, Svecevičius, G., 2002. Comparative analysis of the sensitivity of test-organisms of different phylogenetic level

and live stages to heavy metals. *Environmental and Chemical Physics* (Vilnius, Lithuania) 24(2), 73-78.

- [3] Blinova, I. 2004., Use the freshwater algae and duckweeds for phytoxicity testing. *Environmental Toxicology* 19(4):425-428.
- [4] Montvydienė, D, Lakačauskienė, R, Marčiulionienė, D., 2000. Assessment of toxicity of heavy metal model and natural mixtures for higher plants. *Botanica Lithuanica* (Vilnius, Lithuania) 6(3), 281-297.
- [5] Vosylienė, M.-Z., Marčiulionienė, D., Kazlauskienė, N., Montvydienė, D., Svecevičius, G., 2003. The use of biological test complex for the water toxicity assessment. Lithuanian Society of Metaloecology. Vilnius. p. 16.
- [6] Magone, I., 1989. Bioindication of phytotoxicity of transport emission. In: Kachalova O. L., editor. Bioindication of toxicity of transport emissions in the impact of highway emissions on natural environment. Riga, Latvia: Zinatne. p 108-116.
- [7] Wang, W. 1992., Use of plants for the assessment of environmental contaminants. *Reviews of Environmental Contamination and Toxicology* 126, 88-127.
- [8] Jokšas, et al.,1997. The peculiarity of the transformation and accumulation of sediments and chemical substances in the Druksiai lake. Lithuanian research programme "Nuclear energy and environment" the array of research reports, 1993–1997 I part. Vilnius, 119-225.
- [9] Kazlauskienė, N., Svecevičius, G., Vosylienė, M.-Z., Marčiulionienė, D., Montvydienė, D. 2004. Comparative study on sensitivity of higher plants and fish to heavy fuel oil. *Environmental Toxicology* 19(4), 449-451.
- [10] Wang, W., 1986. Root elongation method for toxicity testing of organic and inorganic pollutants. *Environmental Contamination and Toxicology* 6, 409-414.
- [11] Fargasova, A., 1998. Root growth inhibition, photosynthetic pigments production, and metal accumultation in *Sinapis alba* as the parameters for trace metals effect determination. *Bull Environ Contam Toxicol* 61, 762-769.